

Broad clinical utility built on extensive validation¹⁻⁴

Clinical summary

Our tissue-based comprehensive genomic profiling service for all solid tumours^{1,2,4}

Based on our analytically and clinically validated, FDA-approved comprehensive platform^{1,3}

High concordance with FDA-approved companion diagnostics (CDx)⁴

GENOMIC FINDINGS	POSITIVE PERCENT AGREEMENT (PPA)*	NEGATIVE PERCENT AGREEMENT (NPA)*	COMPARATOR TEST METHOD
<i>EGFR</i> Exon 19 Deletions and L858R Alterations	98.1% (106/108)	99.4% (153/154)	cobas® <i>EGFR</i> Mutation Test v2
<i>EGFR</i> T790M Alterations	98.9% (87/88)	86.1% (93/108)	cobas® <i>EGFR</i> Mutation Test v1 cobas® <i>EGFR</i> Mutation Test v2
<i>ALK</i> Rearrangements	92.9% (78/84)	100% (75/75)	Ventana <i>ALK</i> (D5F3) CDx Assay Vysis <i>ALK</i> Break-Apart FISH Probe Kit
<i>KRAS</i> Alterations	100% (173/173)	100% (154/154)	therascreen® <i>KRAS</i> RGQ PCR Kit
<i>ERBB2</i> (HER2) Amplifications	89.4% (101/113)	98.4% (180/183)	Dako <i>HER2</i> FISH PharmDx® Kit
<i>BRAF</i> V600	99.4% (166/167)	89.6% [†] (121/135)	cobas® <i>BRAF</i> V600 Mutation Test
<i>BRAF</i> V600E	99.3% (149/150)	99.2% (121/122)	
<i>BRAF</i> V600 dinucleotide [‡]	96.3% (26/27)	100% (24/24)	THxID® <i>BRAF</i> kit

- FoundationOne®CDx is concordant with FDA-approved FoundationFocus™ CDx for BRCA 1/2⁴

Cobas is a trademark of Roche Diagnostics Operations, Inc. Therascreen is a trademark of Qiagen. PharmDx is a registered trademark of Dako Denmark A/S. THxID is a registered trademark of bioMérieux. FoundationFocus is a trademark of Foundation Medicine, Inc.

*The reference standard used to calculate positive percent agreement (PPA) and negative percent agreement (NPA) is defined as the consensus calls between the two comparator methods or comparator runs. Agreement calculations solely using consensus calls may overestimate the performance of FoundationOne CDx.

[†]Sensitivity of dinucleotide detection of *BRAF* V600K and V600E was found to be significantly reduced in the cobas® test, in particular for samples in which FoundationOne CDx detected the dinucleotides to be of lower than 40% MAF, leading to low NPA values.

[‡]A study using the THxID™ *BRAF* kit (bioMérieux) was conducted with samples with *BRAF* V600 dinucleotide mutation detected by FoundationOne CDx and *BRAF* V600 negative samples to provide a better evaluation of V600 dinucleotide concordance.

High concordance with externally validated NGS method*⁴

	POSITIVE PERCENT AGREEMENT (PPA)	NEGATIVE PERCENT AGREEMENT (NPA)
All short variants	94.6%	99.9%
Substitutions	96.6%	99.9%
Insertions & deletions	83.4%	99.9%

High concordance with FoundationOne^{®+4}

	POSITIVE PERCENT AGREEMENT (PPA)	NEGATIVE PERCENT AGREEMENT (NPA)
All variants	98.6%	99.9%
All short variants	99.1%	99.9%
Substitutions	99.4%	99.9%
Insertions & deletions	97.0%	99.9%
All copy number variants	94.3%	99.9%
Amplifications	94.0%	99.9%
Losses	94.8%	99.8%
Rearrangements	100.0%	99.9%

- FoundationOne is our pioneering lab-developed test used in over 180,000 patients^{5,6}

*The detection of alterations by FoundationOne CDx was compared to results of an externally validated next-generation sequencing assay (evNGS). Overall there were 157 overlapping genes between the two assays. The comparison between short alterations, including base substitutions and short indels, detected by FoundationOne CDx and the orthogonal method included 188 samples from 46 different tumours. Differences in variants of unknown significance (VUS) alteration calls between the platform were noted, and are expected based on differences in filtering employed by FoundationOne CDx and evNGS. Negative predictive value and positive predictive value were also calculated and were found to be different than percent agreement because the two platforms filter VUS differently. Discordant alterations not related to VUS filtering were primarily caused by deletions with low allelic fraction in homopolymer regions. The FoundationOne CDx variant calling pipeline imposes a filter based on MAF of ≥ 0.10 for indels in homopolymer regions to reduce the likelihood of calling false positives resulting from artifacts introduced by the technology. As such, the difference observed was due to varying filter thresholds between the two platforms.

[†]To support the use of retrospective data generated using FoundationOne, a concordance study was conducted with FoundationOne CDx. This study evaluated a test set of 165 specimens. PPA and NPA between FoundationOne CDx and FoundationOne, using the FoundationOne assay as the reference method, was calculated for all alterations, as well as for alterations binned by type: short variants, copy number alterations (CNAs) and rearrangements. A total of 2325 variants, including 2026 short variants, 266 CNAs and 33 rearrangements were included in the study. NGS, next-generation sequencing.

Broad clinical utility across common cancers

% patients with genomic alterations that the FoundationOne CDx platform is clinically validated to detect

Melanoma⁷⁻¹²

39%-94% patients

BRAF V600E 36% - 75%
BRAF V600K 2% - 19%

Identifies 37% more patients with clinically relevant *BRAF* alterations compared with PCR, IHC and other NGS testing methods⁴⁰

Colorectal cancer¹³⁻²⁰

61%-75% patients

KRAS wildtype 59% - 70%
NRAS wildtype 2% - 5%

Identifies up to 12% more patients with resistance-associated *KRAS* alterations compared with traditional PCR-based methods⁴¹

NSCLC²¹⁻³¹

20%-39% patients

EGFR exon 19 deletions 8% - 17%
EGFR exon 21 L858R 6% - 11%
EGFR exon 20 T790M 1% at diagnosis
EGFR exon 20 T790M 33% - 62% acquired resistance
ALK rearrangements 5% - 8%
BRAF V600E 1% - 2%

Identifies up to 35% and 17% more patients with *ALK* fusions and *EGFR* alterations, respectively, compared with traditional FISH and PCR-based methods^{42,43}

Ovarian³²⁻³⁵

13%-28% patients

BRCA1 8% - 19%
BRCA2 5% - 9%

Identifies patients with *BRCA1/2* alterations who may benefit from PARP inhibitors⁴⁴

Breast³⁶⁻³⁹

13%-23% patients

ERBB2 (HER2) amplification

Identifies patients with *ERBB2* alterations who may benefit from HER2 therapy⁴
Detects drug resistance mechanisms, including various *ESR1* and *RB1* alterations⁴ that may inform patient management regarding hormone therapy and CDK4/6 inhibitors^{44,45}

* FoundationOne CDx does not distinguish between germline and somatic alterations. FISH, fluorescence in situ hybridisation. IHC, immunohistochemistry. NGS, next-generation sequencing. NSCLC, non-small cell lung cancer. PCR, polymerase chain reaction.

Comprehensive analysis of the tumour genome in a single test⁴

Genes with full coding exonic regions included in FoundationOne CDx for the detection of substitutions, insertion-deletions (indels), and copy-number alterations (CNAs).

Assesses 324 cancer-related genes⁴

Genes with full coding exonic regions included in FoundationOne CDx:

For detection of substitutions, insertion-deletions and copy-number alterations

<i>ABL1</i>	<i>ACVR1B</i>	<i>AKT1</i>	<i>AKT2</i>	<i>AKT3</i>	<i>ALK</i>	<i>ALOX12B</i>	<i>AMER1 (FAM123B)</i>	<i>APC</i>
<i>AR</i>	<i>ARAF</i>	<i>ARFRP1</i>	<i>ARID1A</i>	<i>ASXL1</i>	<i>ATM</i>	<i>ATR</i>	<i>ATRAX</i>	<i>AURKA</i>
<i>AURKB</i>	<i>AXIN1</i>	<i>AXL</i>	<i>BAP1</i>	<i>BARD1</i>	<i>BCL2</i>	<i>BCL2L1</i>	<i>BCL2L2</i>	<i>BCL6</i>
<i>BCOR</i>	<i>BCORL1</i>	<i>BRAF</i>	<i>BRCA1</i>	<i>BRCA2</i>	<i>BRD4</i>	<i>BRIP1</i>	<i>BTG1</i>	<i>BTG2</i>
<i>BTK</i>	<i>CT1orf30 (EMSY)</i>	<i>CALR</i>	<i>CARD11</i>	<i>CASP8</i>	<i>CBFB</i>	<i>CBL</i>	<i>CCND1</i>	<i>CCND2</i>
<i>CCND3</i>	<i>CCNE1</i>	<i>CD22</i>	<i>CD274 (PD-L1)</i>	<i>CD70</i>	<i>CD79A</i>	<i>CD79B</i>	<i>CDC73</i>	<i>CDH1</i>
<i>CDK12</i>	<i>CDK4</i>	<i>CDK6</i>	<i>CDK8</i>	<i>CDKN1A</i>	<i>CDKN1B</i>	<i>CDKN2A</i>	<i>CDKN2B</i>	<i>CDKN2C</i>
<i>CEBPA</i>	<i>CHEK1</i>	<i>CHEK2</i>	<i>CIC</i>	<i>CREBBP</i>	<i>CRKL</i>	<i>CSF1R</i>	<i>CSF3R</i>	<i>CTCF</i>
<i>CTNNA1</i>	<i>CTNNB1</i>	<i>CUL3</i>	<i>CUL4A</i>	<i>CXCR4</i>	<i>CYP17A1</i>	<i>DAXX</i>	<i>DDR1</i>	<i>DDR2</i>
<i>DIS3</i>	<i>DNMT3A</i>	<i>DOT1L</i>	<i>EED</i>	<i>EGFR</i>	<i>EP300</i>	<i>EPHA3</i>	<i>EPHB1</i>	<i>EPHB4</i>
<i>ERBB2</i>	<i>ERBB3</i>	<i>ERBB4</i>	<i>ERCC4</i>	<i>ERG</i>	<i>ERRF1</i>	<i>ESR1</i>	<i>EZH2</i>	<i>FAM46C</i>
<i>FANCA</i>	<i>FANCC</i>	<i>FANCG</i>	<i>FANCL</i>	<i>FAS</i>	<i>FBXW7</i>	<i>FGF10</i>	<i>FGF12</i>	<i>FGF14</i>
<i>FGF19</i>	<i>FGF23</i>	<i>FGF3</i>	<i>FGF4</i>	<i>FGF6</i>	<i>FGFR1</i>	<i>FGFR2</i>	<i>FGFR3</i>	<i>FGFR4</i>
<i>FH</i>	<i>FLCN</i>	<i>FLT1</i>	<i>FLT3</i>	<i>FOXL2</i>	<i>FUBP1</i>	<i>GABRA6</i>	<i>GATA3</i>	<i>GATA4</i>
<i>GATA6</i>	<i>GID4 (C17orf39)</i>	<i>GNA11</i>	<i>GNA13</i>	<i>GNAQ</i>	<i>GNAS</i>	<i>GRM3</i>	<i>GSK3B</i>	<i>H3F3A</i>
<i>HDAC1</i>	<i>HGF</i>	<i>HNF1A</i>	<i>HRAS</i>	<i>HSD3B1</i>	<i>ID3</i>	<i>IDH1</i>	<i>IDH2</i>	<i>IGF1R</i>
<i>IKBKE</i>	<i>IKZF1</i>	<i>INPP4B</i>	<i>IRF2</i>	<i>IRF4</i>	<i>IRS2</i>	<i>JAK1</i>	<i>JAK2</i>	<i>JAK3</i>
<i>JUN</i>	<i>KDM5A</i>	<i>KDM5C</i>	<i>KDM6A</i>	<i>KDR</i>	<i>KEAP1</i>	<i>KEL</i>	<i>KIT</i>	<i>KLHL6</i>
<i>KMT2A (MLL)</i>	<i>KMT2D (MLL2)</i>	<i>KRAS</i>	<i>LTK</i>	<i>LYN</i>	<i>MAF</i>	<i>MAP2K1 (MEK1)</i>	<i>MAP2K2 (MEK2)</i>	<i>MAP2K4</i>
<i>MAP3K1</i>	<i>MAP3K13</i>	<i>MAPK1</i>	<i>MCL1</i>	<i>MDM2</i>	<i>MDM4</i>	<i>MED12</i>	<i>MEF2B</i>	<i>MEN1</i>
<i>MERTK</i>	<i>MET</i>	<i>MITF</i>	<i>MKNK1</i>	<i>MLH1</i>	<i>MPL</i>	<i>MRE11A</i>	<i>MSH2</i>	<i>MSH3</i>
<i>MSH6</i>	<i>MST1R</i>	<i>MTAP</i>	<i>MTOR</i>	<i>MUTYH</i>	<i>MYC</i>	<i>MYCL (MYCL1)</i>	<i>MYCN</i>	<i>MYD88</i>
<i>NBN</i>	<i>NF1</i>	<i>NF2</i>	<i>NFE2L2</i>	<i>NFKB1A</i>	<i>NKX2-1</i>	<i>NOTCH1</i>	<i>NOTCH2</i>	<i>NOTCH3</i>
<i>NPM1</i>	<i>NRAS</i>	<i>NT5C2</i>	<i>NTRK1</i>	<i>NTRK2</i>	<i>NTRK3</i>	<i>P2RY8</i>	<i>PALB2</i>	<i>PARK2</i>
<i>PARP1</i>	<i>PARP2</i>	<i>PARP3</i>	<i>PAX5</i>	<i>PBRM1</i>	<i>PDCD1 (PD-1)</i>	<i>PDCD1LG2 (PD-L2)</i>	<i>PDGFRA</i>	<i>PDGFRB</i>
<i>PDK1</i>	<i>PIK3C2B</i>	<i>PIK3C2G</i>	<i>PIK3CA</i>	<i>PIK3CB</i>	<i>PIK3R1</i>	<i>PIM1</i>	<i>PMS2</i>	<i>POLD1</i>
<i>POLE</i>	<i>PPARG</i>	<i>PPP2R1A</i>	<i>PPP2R2A</i>	<i>PRDM1</i>	<i>PRKARIA</i>	<i>PRKCI</i>	<i>PTCH1</i>	<i>PTEN</i>
<i>PTPN11</i>	<i>PTPRO</i>	<i>QKI</i>	<i>RAC1</i>	<i>RAD21</i>	<i>RAD51</i>	<i>RAD51B</i>	<i>RAD51C</i>	<i>RAD51D</i>
<i>RAD52</i>	<i>RAD54L</i>	<i>RAF1</i>	<i>RARA</i>	<i>RB1</i>	<i>RBM10</i>	<i>REL</i>	<i>RET</i>	<i>RICTOR</i>
<i>RNF43</i>	<i>ROSI</i>	<i>RPTOR</i>	<i>SDHA</i>	<i>SDHB</i>	<i>SDHC</i>	<i>SDHD</i>	<i>SETD2</i>	<i>SF3B1</i>
<i>SGK1</i>	<i>SMAD2</i>	<i>SMAD4</i>	<i>SMARCA4</i>	<i>SMARCB1</i>	<i>SMO</i>	<i>SNCAIP</i>	<i>SOCS1</i>	<i>SOX2</i>
<i>SOX9</i>	<i>SPEN</i>	<i>SPOP</i>	<i>SRC</i>	<i>STAG2</i>	<i>STAT3</i>	<i>STK11</i>	<i>SUFU</i>	<i>SYK</i>
<i>TBX3</i>	<i>TEK</i>	<i>TET2</i>	<i>TGFBR2</i>	<i>TIPARP</i>	<i>TNFAIP3</i>	<i>TNFRSF14</i>	<i>TP53</i>	<i>TSC1</i>
<i>TSC2</i>	<i>TYRO3</i>	<i>U2AF1</i>	<i>VEGFA</i>	<i>VHL</i>	<i>WHSC1 (MMSET)</i>	<i>WHSC1L1</i>	<i>WT1</i>	<i>XPO1</i>
<i>XRCC2</i>	<i>ZNF217</i>	<i>ZNF703</i>						

Select rearrangements:

Genes with select intronic regions for the detection of gene rearrangements, one gene with a promoter region and one non-coding RNA gene.

<i>ALK</i>	<i>BCL2</i>	<i>BCR</i>	<i>BRAF</i>	<i>BRCA1</i>	<i>BRCA2</i>	<i>CD74</i>	<i>EGFR</i>	<i>ETV4</i>
<i>ETV5</i>	<i>ETV6</i>	<i>EWSR1</i>	<i>EZR</i>	<i>FGFR1</i>	<i>FGFR2</i>	<i>FGFR3</i>	<i>KIT</i>	<i>KM2A (MLL)</i>
<i>MSH2</i>	<i>MYB</i>	<i>MYC</i>	<i>NOTCH2</i>	<i>NTRK1</i>	<i>NTRK2</i>	<i>NUTM1</i>	<i>PDGFRA</i>	<i>RAF1</i>
<i>RARA</i>	<i>RET</i>	<i>ROSI</i>	<i>RSPO2</i>	<i>SDC4</i>	<i>SLC34A2</i>	<i>TERC*</i>	<i>TERT (promoter only)†</i>	
<i>TMPRSS2</i>								

In the same test reports:⁴

- Tumour mutational burden
- Microsatellite instability

* *TERC* is a non-coding RNA gene. † *TERT* is a gene with a promoter region.

Broad clinical utility built on extensive validation¹⁻⁴



Extensive validation¹⁻³

- » Based on our analytically and clinically validated, FDA-approved comprehensive platform^{1,3}
- » High concordance with FDA-approved companion diagnostics and with NGS methods⁴



Broad clinical utility

- » Clinically relevant results across common cancers: colorectal cancer, melanoma, NSCLC, ovarian and breast⁷⁻³⁹

324
GENES
TMB+MSI

Goes beyond common genomic alterations⁴

- » Assesses 324 cancer-relevant genes⁴
- » Reports TMB and MSI⁴

Intended use statement

FoundationOne®CDx is a next generation sequencing based *in vitro* diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including tumour mutational burden (TMB) and microsatellite instability (MSI) using DNA isolated from formalin-fixed paraffin embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, FoundationOne CDx is intended to provide tumour mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

For more information, please contact your Roche representative

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References

1. FoundationOne®CDx FDA Approval, 2017. Available at: https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019a.pdf (Accessed August 2018);
2. FoundationOne®CDx FDA Approval Press Release, 2017. Available at: <https://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm587273.htm> (Accessed August 2018);
3. FoundationOne®CDx clinical validation, 2017. Available at: <http://www.foundationmedicine.com/genomic-testing/foundation-one-cdx> (Accessed August 2018);
4. FoundationOne®CDx Technical Specifications 2018. Available at: www.rochefoundationmedicine.com/flcdxtech;
5. Frampton GM *et al. Nat Biotechnol* 2013; 31:1023-1031;
6. Foundation Insights. Available at: <https://www.foundationmedicine.com/insights-and-trials/foundation-insights> (Accessed August 2018);
7. Greaves WO *et al. J Mol Diagn* 2013; 15:220-226;
8. Davies H *et al. Nature* 2002; 417:949-954;
9. Hodis E *et al. Cell* 2012; 150:251-263;
10. Menzies AM *et al. Clin Cancer Res* 2012; 18:3242-3249;
11. Colombino M *et al. J Clin Oncol* 2012; 30:2522-2529;
12. Long GV *et al. J Clin Oncol* 2011; 29:1239-1246;
13. Roth AD *et al. J Clin Oncol* 2010; 28:466-474;
14. Amado RG *et al. J Clin Oncol* 2008; 26:1626-1634;
15. Douillard JY *et al. N Engl J Med* 2013; 369:1023-1034;
16. Heinemann V *et al. Lancet Oncol* 2014; 15:1065-1075;
17. Price TJ *et al. Br J Cancer* 2015; 112:963-970;
18. De Roock W *et al. Lancet Oncol* 2010; 11:753-762;
19. Vaughn CP *et al. Genes Chromosomes Cancer* 2011; 50:301-312;
20. Peeters M *et al. Clin Cancer Res* 2013; 19:1902-1912;
21. Vanderlaen PA *et al. Lung Cancer* 2018; 116:90-95;
22. Kris MG *et al. JAMA* 2014; 311:1998-2006;
23. D'Angelo SP *et al. J Clin Oncol* 2011; 29:2066-2070;
24. Esteban E *et al. Cancer Epidemiol* 2015; 39:291-297;
25. Han B *et al. Lung Cancer* 2017; 113:37-44;
26. Barlesi F *et al. Lancet* 2016; 387:1415-1426;
27. Hata A *et al. Cancer* 2013; 119: 325-4332;
28. Tanaka K *et al. Oncotarget* 2017; 8:68123-68130;
29. Sequist LV *et al. Sci Transl Med* 2011; 3:75ra26;
30. Oxnard GR *et al. Clin Cancer Res* 2011; 17:1616-1622;
31. Paik PK *et al. J Clin Oncol* 2011; 29:2046-2051;
32. Yang D *et al. JAMA* 2011; 306:1557-1565;
33. Cancer Genome Atlas Research Network Nature 2011; 474:609-615;
34. Zhang S *et al. Gynecol Oncol* 2011; 121:353-357;
35. Pennington KP *et al. Clin Cancer Res* 2014; 20:764-775;
36. Cancer Genome Atlas Network Nature 2012; 490:61-70;
37. Owens MA *et al. Clin Breast Cancer* 2004; 5:63-69;
38. Chmielecki J *et al. Oncologist* 2015; 20:7-12;
39. Bartlett JM *et al. J Pathol* 2001; 195:422-428;
40. Boussemart *et al. Presented at ESMO (European Society for Medical Oncology) 2017, Madrid, Spain: Poster no. 1234;*
41. Rankin A *et al. Oncologist* 2016; 21:1306-1314;
42. Ali SM *et al. Oncologist* 2016; 21:762-770;
43. Schrock AB *et al. Clin Cancer Res* 2016; 22:3281-3285.
44. Angus L *et al. Cancer Treatment Reviews* 2017; 52:33-40;
45. Condorelli R *et al. Ann Oncol* 2018; 29:640-645.

